# Full-length V(D)J immune repertoire sequencing (IR-Seq) on the NextSeq™ 1000 and NextSeq 2000 Systems

- More data with higher-quality reads allows discovery of more clonotypes

- More samples per run for flexible experimental designs to meet research budgets

# Immune repertoire biology

The adaptive immune system, made up of B lymphocytes and T lymphocytes, provides highly specific protection from pathogens through an evolutionarily refined process that produces a massively diverse repertoire of antigen receptors.[1] When an antigen receptor recognizes a pathogen, a series of events takes place to amplify that receptor and recruit additional immune cells to eliminate the infection. This process for generating receptor diversity is known as V(D)J recombination. In V(D)J recombination, variable (V), diversity (D), and joining (J) segments are stochastically recombined into functional B-cell receptors (BCRs) and T-cell receptors (TCRs). Genes for BCRs and TCRs occur at different sites in the genome as arrays of variable V, D, and J segments, as well as constant regions that make up the rest of the receptor structure (Figure 1 and Figure 2). By randomly recombining V, D, and J segments from a small number of genes into mRNA transcripts, it is estimated that it is possible to generate $10^{18}$ unique BCR sequences[2,3] and $10^{12}$ unique TCR sequences.[4,5]

The quantifiable immune repertoire of BCR and TCR sequences is dynamic in response to infections, autoimmune disorders (eg, lupus, rheumatoid arthritis,
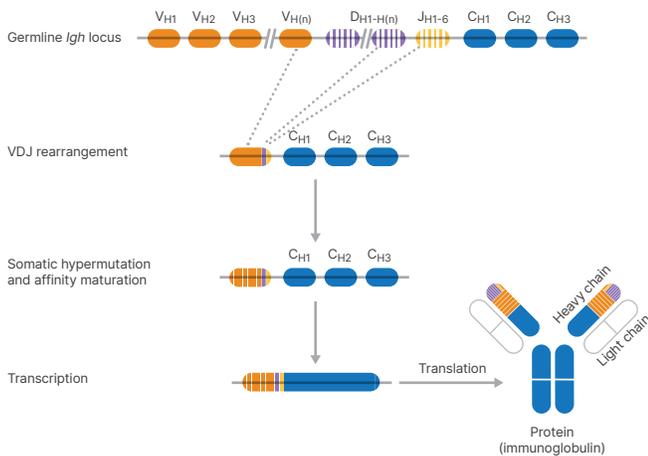


Figure 1: BCR generation through V(D)J recombination process— Genes for BCR receptors occur at multiple sites in the genome. The genes contain a large number of V domains and a smaller number of D and J domains, as well as constant C domains. Stochastic recombination of some of the V(D)J domains results in a highly diverse population of immunoglobulin receptors.
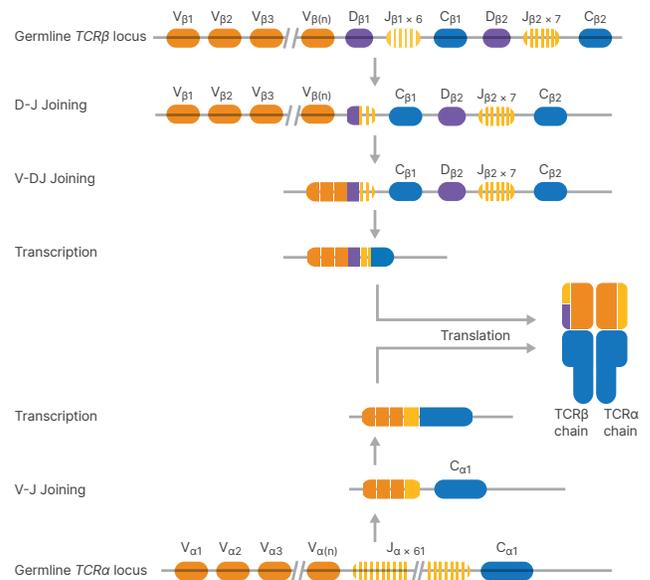


Figure 2: TCR generation through V(D)J recombination process— TCRs are heterodimers transcribed from two different genomic loci. The TCRβ gene contains several V, D, J, and constant domains while the TCRα contains V, J, and constant domains. Stochastic recombination of V, D, and J domains results in a highly diverse population of TCRs.

multiple sclerosis), and cancers. During an immune response, the repertoire of circulating antigen receptors shifts from a diverse pool to one that is dominated by on or a few expanded clones (Figure 3). Therefore, understanding the makeup of the repertoire is highly informative for what is happening in the disease process. The incredible diversity of the immune repertoire means that analysis requires high sequencing depth. In addition, the ability to accurately assemble the full-length recombined V(D)J genes plus partial constant region containing isotype-determining information that ranges from 400 bp to 600 bp[6] requires a platform capable of 2 × 300 paired-end reads to form a single, gapless contig for the variable region sequence reconstruction.

The availability of 600-cycle kits makes the NextSeq 1000 and NextSeq 2000 Systems an excellent choice for labs looking for a powerful benchtop sequencer for IR-Seq and other NGS applications. The NextSeq 1000/2000 P1 Reagents (600 cycles) (Illumina, Catalog no. 20075294) generates up to 60 Gb of high-quality data with 100M reads and the NextSeq 1000/2000 P2 300M Reagents (600 cycles) (Illumina, Catalog no. 20075295) generates up to 180 Gb of high-quality data with 300M reads. In

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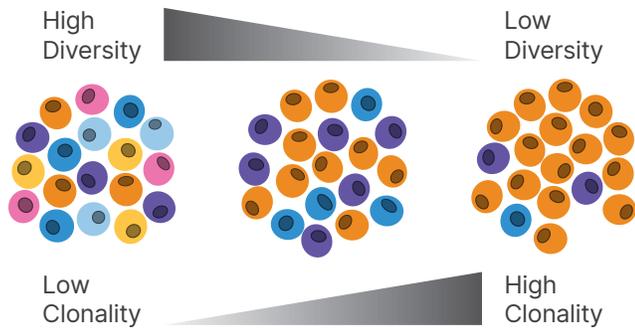
Figure 3: Adaptive immune response—The adaptive immune system relies on a highly diverse population of cells expressing unique BCRs and TCRs. In the absence of an antigen response, the repertoire shows high diversity and low clonality of receptor types. In the presence of a recognized antigen, cells with activated receptors are expande,d resulting in low diversity in the repertoire and high clonality of the active cells.

comparison, the MiSeq™ Reagent Kit v3 (600-cycle) (Illumina, Catalog number MS-102-3003) generates up to 15 Gb of high-quality data with 22.5M reads.

# IR-Seq library preparation

The choice of an immune repertoire library solution depends on various factors such as input (ie, DNA, bulk RNA, or single-cell RNA), methodology (ie, rapid amplification of cDNA ends or multiplex PCR), regions sequenced (ie, complementarity determining region (CDR3) or full-length), and the chain type (ie, TCR or BCR).[7] Multiple library prep solutions and protocols for immune repertoire studies are available and several have been tested on the NextSeq 2000 System (Table 1). Many of these options require or benefit from 2 × 300 paired-end sequencing. The NextSeq 1000/2000 Reagents (600 cycles) represents an advanced alternative to the MiSeq Reagent Kit v3 (600-cycle) for researchers looking to scale their experiments.

Table 1: Tested third-party library prep kit for IR-Seq applications[a]

| Resolution | Method | Provider | Kit name (Catalog no.) | Targeted receptors | Sequencing read length recommendation | Bioinformatics secondary analysis |
|---|---|---|---|---|---|---|
| Bulk RNA-Seq | CDR3/full-length | New England Biolabs | NEBNext Immune Sequencing Kit (Human) (E6320S, E6320L) | BCR, TCR or BCR + TCR | 2 × 300 bp | Open Source pRESTO Tools |
| Bulk RNA-Seq | CDR3/full-length | New England Biolabs | NEBNext Immune Sequencing Kit (Mouse) (E6330S, E6330L) | BCR, TCR or BCR + TCR | 2 × 300 bp | Open Source pRESTO Tools |
| Bulk RNA-Seq | CDR3/full-length | QIAGEN | QIAseq Immune Repertoire RNA Library Kit (333705) | TCR | 2 × 300 bp | QIAGEN GeneGlobe pipeline |
| Bulk RNA-Seq | CDR3/full-length | Takara | SMART-Seq Human TCR (with UMIs) (634780, 634781, 634779) | TCR | 2 × 300 bp | Cogent NGS Immune Profiler Software |
| Bulk RNA-Seq | CDR3/full-length | Takara | SMART-Seq Human BCR (with UMIs) (634777, 634778, 634776) | BCR | 2 × 300 bp | Cogent NGS Immune Profiler Software |
| Single cell | CDR3/full-length | BD Bio | BD Rhapsody TCR/BCR Multiomic Assay Kit (665828, 665829) | BCR, TCR or BCR + TCR | 85 × 215 bp[b] or 2 × 300 bp | BD Rhapsody Analysis Pipeline |

a. Represented libraries use unique molecular index (UMI) technology, providing error correction, deduplication, and high confidence reads coming out of the pipeline.
b. Run configurations for single cell are compatible with a smaller 300-cycles kit cartridge.

## IR-Seq performance

IR-Seq libraries typically have large fragment sizes (600–800 bp) and a limited capability for multiplexing due to read depth requirements. The The NextSeq 1000/2000 P1 Reagents (600 cycles) can generate 100M reads, approximately 4× more data than the MiSeq System using the MiSeq Reagent Kit v3 (600-cycle). The NextSeq 1000/2000 P2 300M Reagents (600 cycles) generates an impressive 300M reads, or 12× more data versus the MiSeq System using the MiSeq Reagent Kit v3 (600-cycle). This substantial data capacity on the NextSeq 1000 and NextSeq 2000 Systems adds flexibility for increased read depth over the MiSeq System and for multiplexing of IR-Seq libraries (Table 2).

The The NextSeq 1000/2000 P1 Reagents (600 cycles) and NextSeq 1000/2000 P2 300M Reagents (600 cycles) on the NextSeq 2000 system also produce a higher percentage of reads with quality scores ≥ Q30 compared to the MiSeq Reagent Kit v3 (600-cycle) on the MiSeq System. The improved Q30 scores across the length of the reads allow for better secondary analysis results and better clonotype identification (Table 2).

Finally, sequencing on the The NextSeq 1000/2000 P1 Reagents (600 cycles) is significantly faster than the MiSeq System. The time savings could allow labs to complete three sequencing runs per week with the NextSeq 1000/2000 P1 Reagents (600 cycles), with more data per run, compared to two runs per week using the MiSeq Reagent Kit v3 (600-cycle) (Table 2).

Sequencing data generated with the NextSeq 1000 and NextSeq 2000 Systems can be output in standardized file formats that are compatible with an extensive ecosystem of commercial and open-source bioinformatic analysis tools. Each library prep provider offers their own custom analysis. Also, third-party dedicated tools are available for immune repertoire analysis.

## Summary

The state of the adaptive immune system is directly tied to the health of an individual. Therefore, understanding the sequence composition of the immune repertoire provides researchers with a wealth of information about the health of a person, including how they are responding to an infection, the health of their immune system, and even the progress of certain cancers.

NextSeq 1000 and NextSeq 2000 Systems are compatible with a wide range of library preparation kits and analysis software from Illumina as well as third-party suppliers. With the available 600-cycle kits, these systems are excellent for complex IR-Seq applications and provide the reads necessary for a detailed view of the immune repertoire and the capacity for multiplexing of samples. Different flow cell configurations allow researchers to adjust read depth and sample numbers to match their experimental needs. The NextSeq 1000 and NextSeq 2000 Sequencing Systems offer extensive cross-application flexibility, enabling researchers to transition easily between sequencing projects.

Table 2: NextSeq 1000, NextSeq 2000, and MiSeq Systems 600-cycle kit performance

| | MiSeq Reagent Kit v3 (600-cycle) | NextSeq 1000/2000 P1 Reagents (600 cycles) | NextSeq 1000/2000 P2 300M Reagents (600 cycles) |
|---|---|---|---|
| Quality scores (percentage of bases ≥ Q30) | ≥ 70% | ≥ 80% | ≥ 80% |
| Maximum total read pairs per run | 25M | 100M | 300M |
| Maximum sample read pairs per run[a] | 22.5M | 90M | 270M |
| Sample Multiplexing to reach 5M reads[a] | 4 | 18 | 54 |
| Sample Multiplexing to reach 25M reads[a] | – | 3 | 10 |
| Run times | 55 hr | 34 hr | 44 hr |

a. Maximum sample reads when using 10% (v/v) PhiX library addition for sequence diversity.

## Learn more

NextSeq 1000 and NextSeq 2000 system

Immune-Related Genetic Variation

QIAGEN QIAseq Immune Repertoire RNA Library Kit data on the NextSeq 2000 600-cycles kit

New England Biolabs NEBNext Immune Sequencing Kit data now available on the new NextSeq 2000 600-cycles kit

Takara SMARTer Human BCR data now available on new 600-cycles kit on the NextSeq 2000 System

## References

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